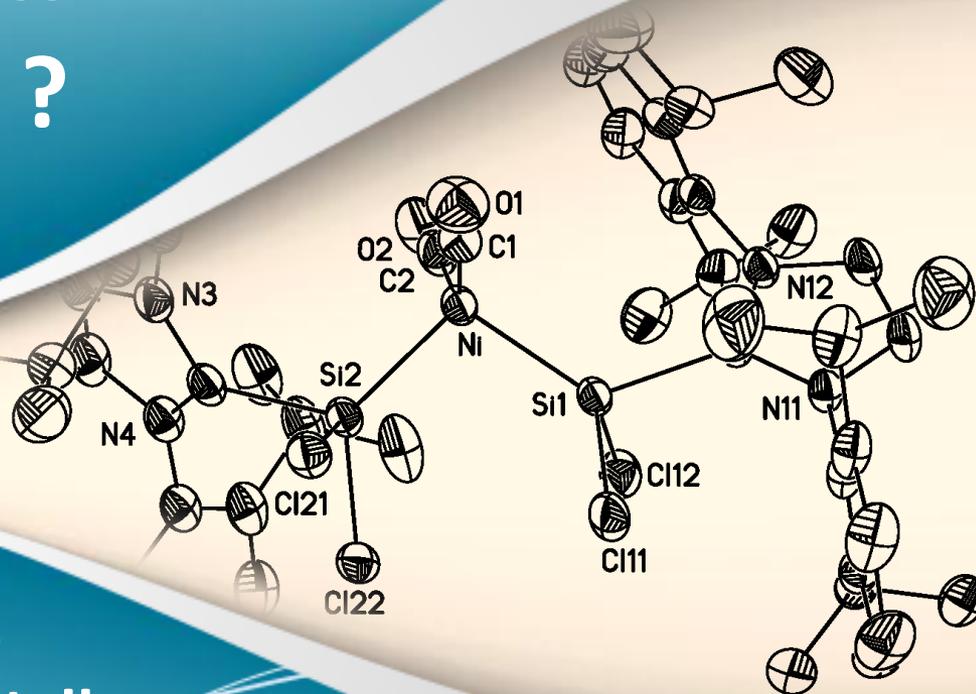
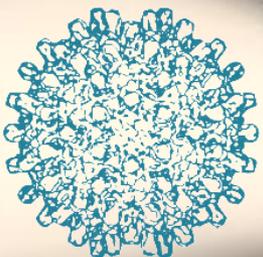
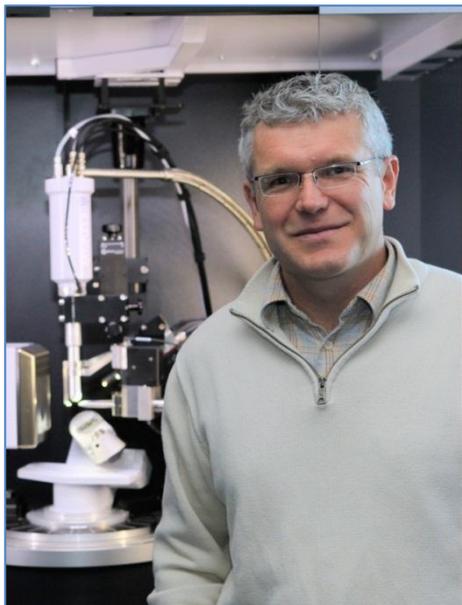


Bruker Webinar What's new in SHELXL-2013 ?



Andrea Thorn
12th September, 2013
Michael Ruf & Bruce C. Noll

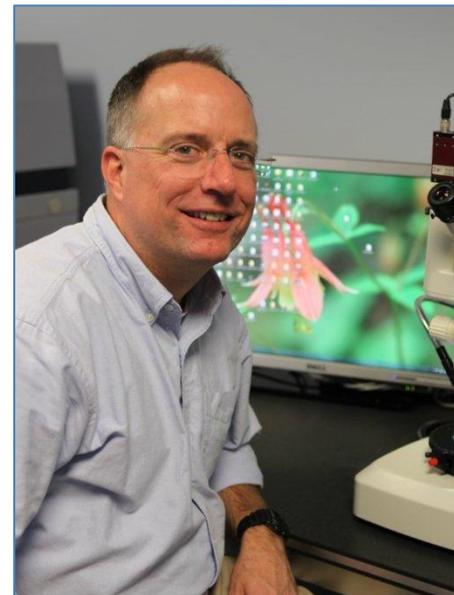
Welcome



Dr. Michael Ruf
Product Manager, SC-XRD
Bruker AXS Inc.
Madison, WI, USA



Andrea Thorn
Crystallographic Computing Group
Medical Research Council –
Laboratory of Molecular Biology
Cambridge, UK



Bruce C. Noll, Ph.D.
Sr. Applications Scientist, SC-XRD
Bruker AXS Inc.
Madison, WI, USA

Introduction: SHELXL_2013

- SHELXL_2013 is a program for the refinement of small and macromolecular crystal structures using single-crystal diffraction data from X-rays or neutrons.
- Stand-alone executables (no libraries, extra files or environment variables required).
- Compatible with all modern versions of Linux, Windows and MacOSX.
- Free for academic use.
- Compatible with SHELX76 and SHELXL-97.

Overview

Introduction

New features:

- Multiprocessor capability
- ADP restraints and constraints
- Neutron data refinement

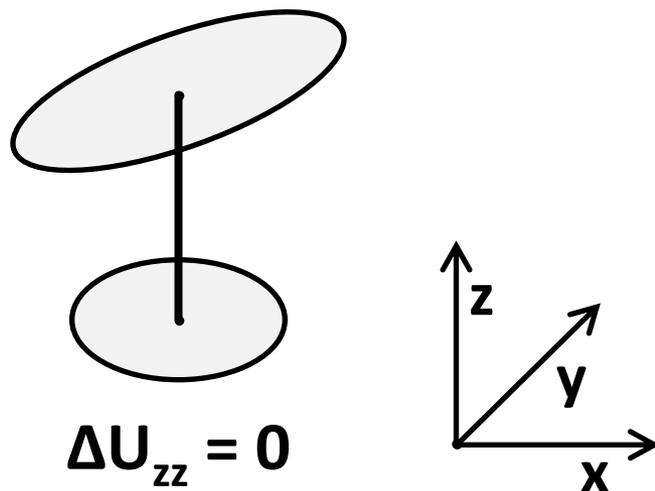
Remarks:

- Estimated standard deviations
- Absolute structure determination
- CIF matters
- Input and output

Rigid-bond restraints

Hirshfeld criterion for small molecule structure validation:

The mean-square displacement amplitudes of bonded atoms are equal in the direction of the bond joining them.



Rigid-bond restraints

Rollett (1970): Rigid-bond restraints

DELU

- Improvement of the data/parameter ratio; enables **anisotropic refinement at relatively good resolution.**
- Standard deviation usually 0.01 - 0.001 Å²
- Provides about one restraint per atom.
- If applied to 1,3-distances as well:
about two restraints per atom

Similar-U restraints

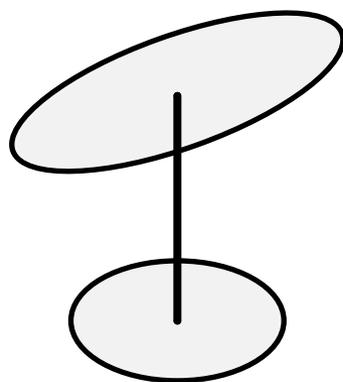
Similar U restraints: All six U_{ij} values of two bonded atoms should be as equal as possible.

SIMU

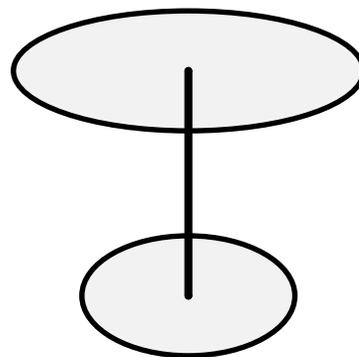
- Large estimated standard deviations
- Much less justified by theory and experimental evidence

Enhanced rigid-bond restraints

If a bond between two atoms is really rigid, the relative motion of the two atoms should be perpendicular to the bond!

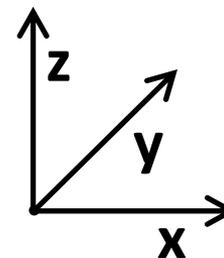


$$\Delta U_{zz} = 0$$



$$\Delta U_{zz} = 0$$

$$\Delta U_{yz} = 0; \Delta U_{xz} = 0$$



Three restraints per atom; if applied to 1,3-distances:
Approximately 6 restraints per atom!

Enhanced rigid-bond restraints

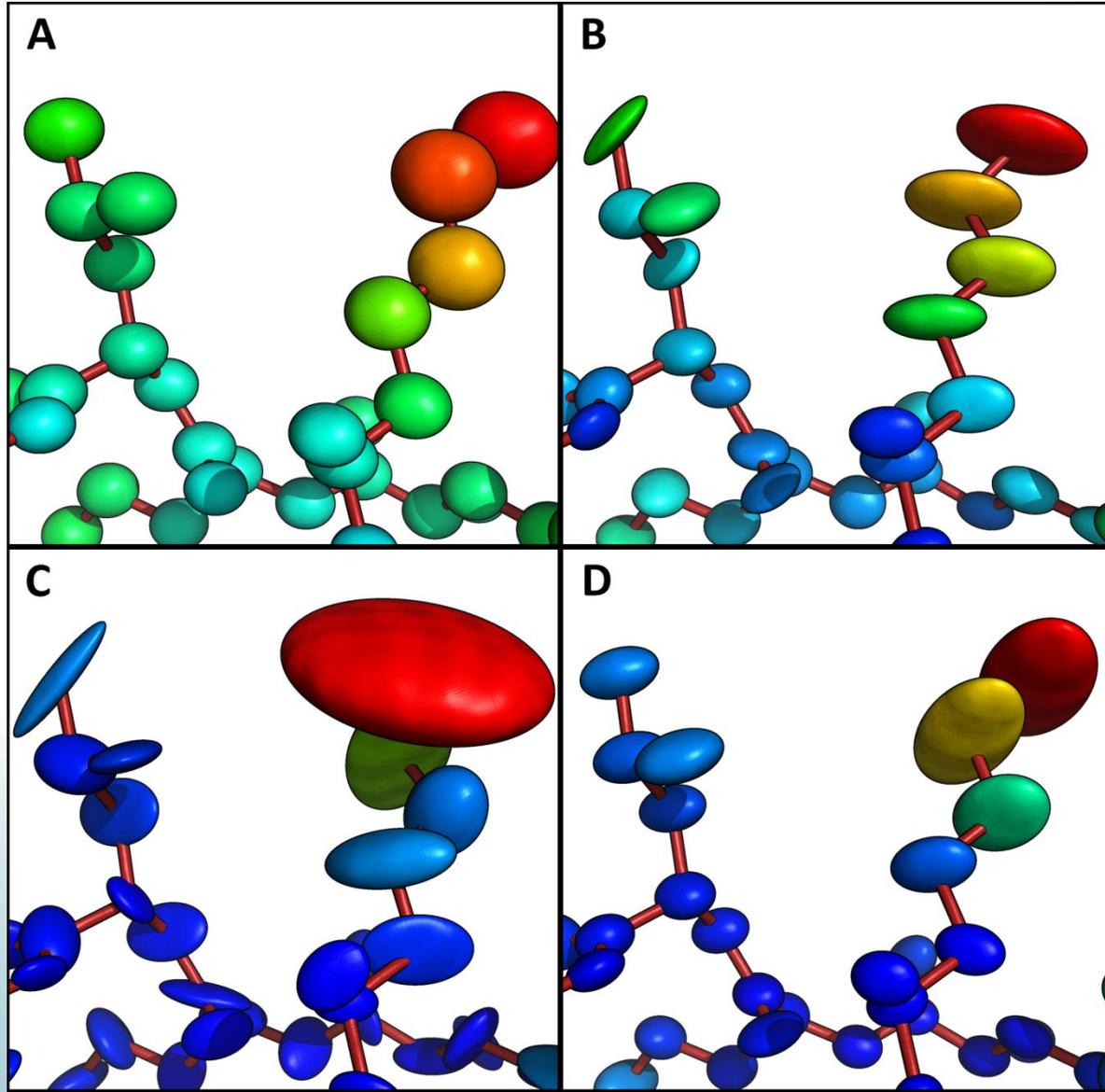
Enhanced rigid-bond restraint in SHELXL

RIGU

Three restraints per atom; if applied to 1,3-distances:
Approximately 6 restraints per atom!

The **new rigid-bond restraints** are an enhancement to the established rigid-bond restraints. However, they cannot account for occupancies dropping along side chains, but may expose such defects.

Tests



PDB 1us0
at 1.7 Å

XNPD constraint

XNPD [-0.001]

- New constraint
- Lower bound for the eigenvalues of the U_{ij} tensor of all anisotropic atoms –
- or the U of isotropic atoms
- The slightly negative default ensures that NPD reports but the refinement does not blow up.

Hydrogen atoms and neutron diffraction

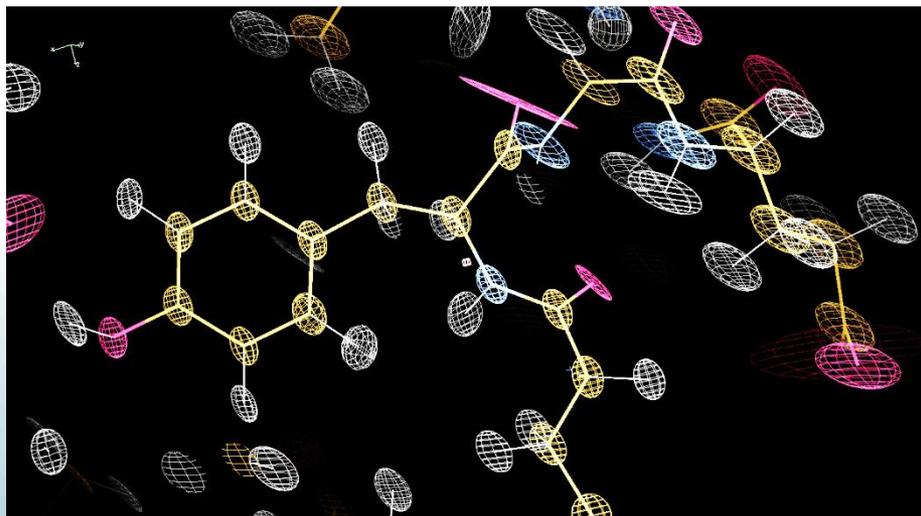
NEUT

To be inserted before **SFAC**: neutron scattering factors default **HFIX/AFIX** bond lengths are changed

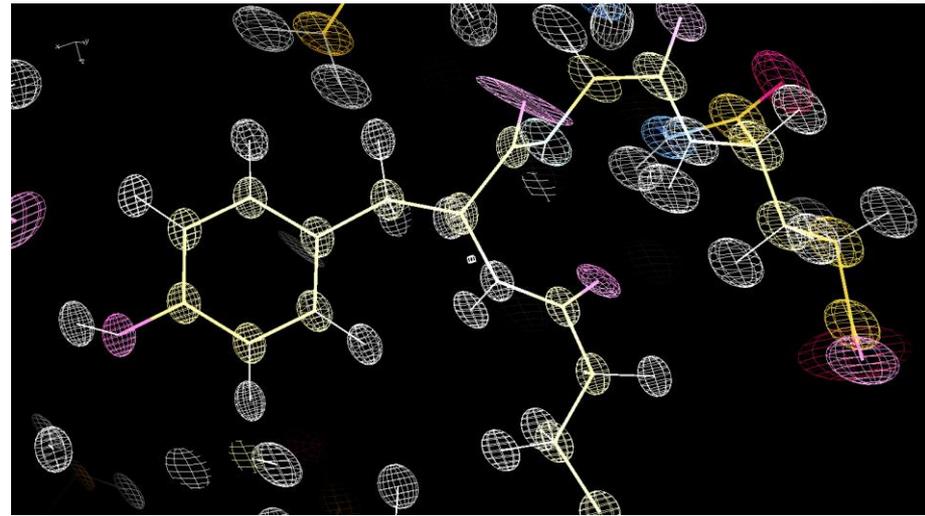
- H/D mostly treated as normal atoms, except for chiral volume restraints (**CHIV**) and riding model (**AFIX**).
- **DFIX** and **SADI** distance restraints work better than the H or D riding model for neutron data.
- If **CHIV** is applied to an atom that is bonded to exactly three non-H/D atoms, bonds to H/D are ignored.
 - Thus the same **CHIV** restraints can be used as for X-rays.
- For atoms bonded to 3 atoms including H/D, all are used.

Anisotropic hydrogens?!

RIGU works well for H/D when refining against neutron data, despite the poor data to parameter ratio, as shown below for myoglobin 1.5Å neutron data (PDB 1L2K).



RIGU + XNDP 0.001



RIGU + XNDP 0.001 + ISOR 0.04 0.04

With RIGU alone, atoms move at right angles to the bonds but ORTEPs are unreasonable. Correction with **ISOR**.

Estimated standard uncertainties

Standard uncertainties obtained by least-squares are proportional to

$$(N-P)^{-1/2}$$

N number of observations

P number of parameters

SHELXL _2013 implements a proposal by Ton Spek, that *N* should always be set to the number of unique reflections.

Estimated standard uncertainties

Consequences:

1. The e.s.d.s no longer depend on how data were merged. Unmerged data should be read into SHELXL for more complete statistics. **MERG 4** may still be used for macromolecules.
2. For HKLF5 twin refinements, ALL data can be used.
3. Some standard uncertainties may be slightly greater than before. They were probably underestimated.
4. The third L.S. parameter is no longer required (except for squeezed structures).

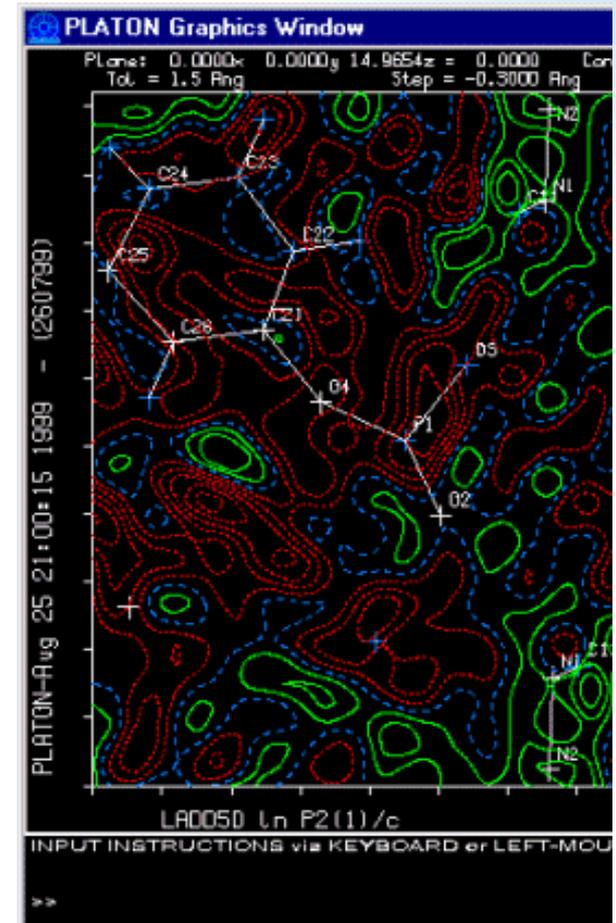
Additional structure factor contribution

ABIN

Input of A and B parts of a calculated structure factor:

- e.g. a *squeezed* region of the cell
- or a solvent mask contribution of a macromolecule

This is especially designed to be compatible with twinned refinements and with Ton Spek's PLATON.



Absolute structure determination

Absolute structure from weak anomalous signal:

SHELXL now calculates the Flack parameter by the Parsons' quotient method at the end of the refinement and outputs the results to the console, *.lst* and *.cif*, for example:

```
Flack x = 0.012 (14) from 271 selected quotients (Parsons' method)
```

Parsons, Flack & Wagner: Post-refinement against Friedel differences or quotients $Q = (I_+ - I_-) / (I_+ + I_-)$ gives much lower e.s.d.s than including the Flack parameter in the full-matrix refinement. The quotients cancel systematic errors (such as bonding electron density) that affect I_+ and I_- equally, and in SHELXL-2013 even give good results for twins.

CIF matters

SHELX-2013 contains extensive changes to its **CIF output** and should be fully compatible with core-CIF version 2.4.3.

The SHELXL-97 definition of *atom site multiplicity* was different to the IUCr definition. This has been fixed by calling the SHELX number *order* instead. Problems may still arise for disordered atoms or groups close to but not exactly on special positions, which is why SHELX never uses this concept (except for writing it to the CIF file).

Improving CIF as an archive format

The *.hkl* file (HKLF 3, 4 or 5 format) and *.res* file are now embedded into the *.cif* output file (with checksums). This makes it possible to repeat any refinement exactly, and discourages cosmetic editing.

- The program **ShredCIF** is provided for extracting the embedded files and verifying their checksums.

SHELXL now deduces the space group name and Hall symbol from LATT and SYMM, but only for 249 common settings. The space group is also written to a COOT-compatible *.pdb* file.

Further new features

- **HFIX** sometimes generates a few bad connectivity error messages when first applied. These are now all printed before aborting. They may then be fixed by **HFIX 0** (before the current **HFIXs**) or **FREE** or **BIND** instructions.
- **HTAB** without atom names now writes the appropriate **EQIV** and full **HTAB** instructions to the end of the *.res* file so that they can be used in the next refinement job. Non-classical C-H..O hydrogen bonds are also generated.
- **SADI** without atom names expands all **SAME** instructions into **SADI** and adds them at the end of the *.res* file.

ACKNOWLEDGEMENTS

George Sheldrick

**The Crystallographic Computing group at the LMB –
Garib Murshudov, Rob Nicholls, Paul Emsley, Fei Long**

Richard Cooper

The programs are **free for academic use:**

<http://shelx.uni-ac.gwdg.de/SHELX/>



MRC

Laboratory of
Molecular Biology

Standard deviations

Usage as restraint:

$$\frac{\sqrt{p^2 + U_{eq,A} + U_{eq,B}} \sigma}{p} \quad \frac{\sqrt{p^2 + U_{eq,A} + U_{eq,B}} \sigma d}{p}$$

A and B are the two atoms

U_{eq} equivalent isotropic displacement parameter

d is the distance between atoms A and B

σ and p are user-supplied parameters

(0.004 Å² and $p = 0.5$ gave good results)

ADPs are not allowed to be non-positive definite.

Verification at high resolution

- Refinement of a cyclic hexapeptide
- 0.382 Å; 100 K
- Cut to 0.84 Å

	IAM_{0.84} 1,2	IAM 1,2	XD 1,2	IAM_{0.84} 1,3	IAM 1,3	XD 1,3
r.m.s. ΔU_{ZZ} [Å²]	0.00165	0.00034	0.00031	0.00188	0.00074	0.00076
r.m.s. $\Delta U_{XZ\&YZ}$ [Å²]	0.00258	0.00180	0.00183	0.00301	0.00242	0.00240
r.m.s. ΔU_{XY} [Å²]	0.00406	0.00304	0.00304	0.00389	0.00386	0.00393
r.m.s. $\Delta U_{XX\&YY}$ [Å²]	0.00665	0.00605	0.00600	0.00788	0.00761	0.00767

IAM = Independent atom refinement

XD = XD multipole refinement

Test set

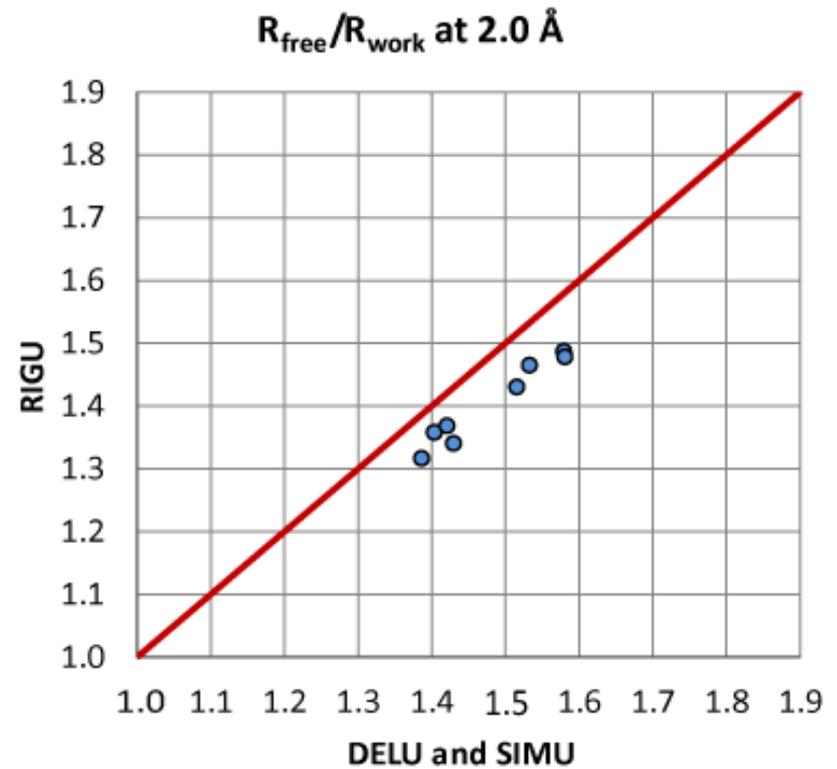
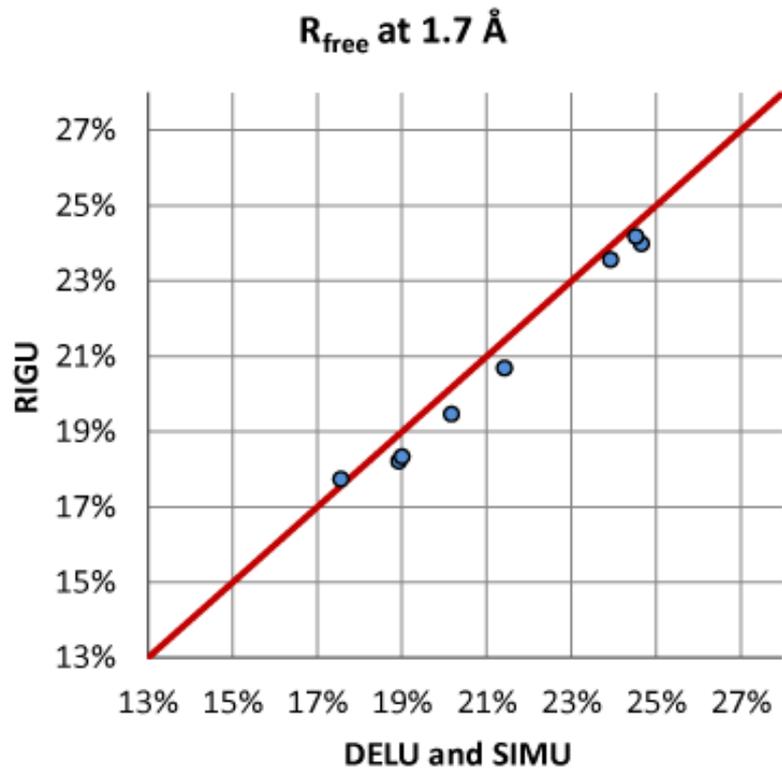
Test set: **Eight well-refined protein structures** *

- 0.7 - 1.2 Å
- 55 - 331 residues
- minor components of disordered residues removed
- water molecules refined isotropically
- structures 'shaken' by random shifts (mean of 0.5 Å)

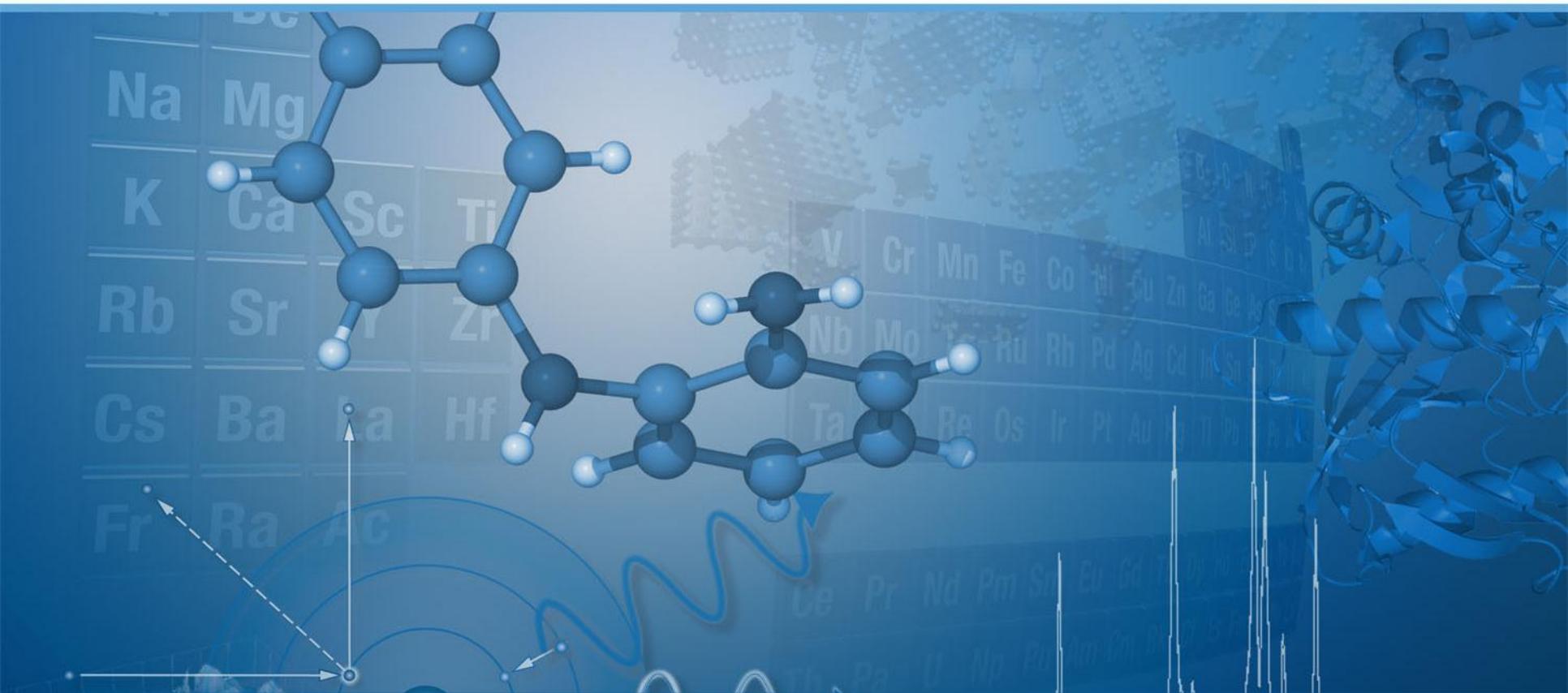
* 1b0y, 1lu0, 1ok0, 1rqw, 1us0, 2cm5, 2fdn and 2vb1

Tests

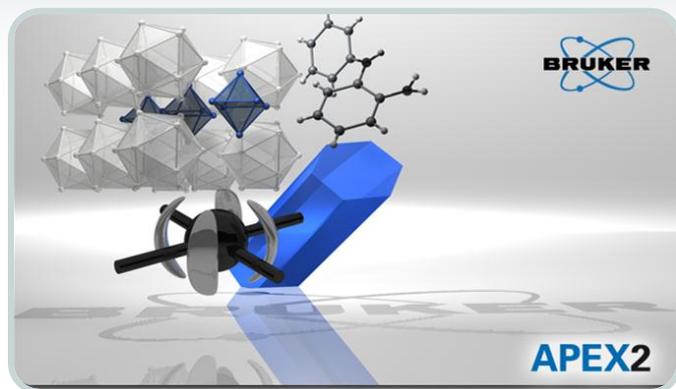
1.7 Å resolution



Using SHELXL-2013 with APEX2



APEX2 and SHELXL-2013



```
+++++  
+ SHELXL - CRYSTAL STRUCTURE REFINEMENT - MULTI-CPU VERSION +  
+ Copyright(C) George M. Sheldrick 1993-2013 Version 2013/4 +  
+ twin5 started at 16:12:13 on 21-Aug-2013 +  
+++++
```

Command line parameters: twin5 -a50000 -b3000 -c624 -t4

-a sets the approximate maximum number of atoms including hydrogens.
-b sets the maximum number of full-matrix parameters (leave unchanged for CGLS). For example -b9000 allows refinement of 1000 anisotropic atoms or 3000 with BLOC 1. For a 32-bit version, -b times the square root of the number of threads should not exceed about 65500. -c sets the reflection buffer size. This depends on the CPU cache size but will rarely need changing. -t sets the number of threads, otherwise the multi-CPU version sets this equal to the number of available CPUs. For optimal performance on systems with hyperthreading, usually the hyperthreading should be switched off or -t used to halve the number of threads; e.g. -t4 rather than -t8 for an Intel i7 processor.

Running 4 threads on 4 processors

- SHELXL is engine underlying refinement in APEX2
- Structure Refinement plug-in allows easy interaction with SHELXL-2013
- Refinement instructions handled directly
- New CIF requirements met during refinement

Refinement



SHELXL
ANSC
RIGU ANSR
ABIN
PRIG NEUT
WIGL
XNPD
TWST

- All new commands can be used within Structure Refinement plug-in

The Structure Refinement Plug-In



APEX2 v2013.8-RC2 - User: Bruce Noll (bnoll) - Sample: sucrose_stress - Temporary license (115 days remaining) - [Structure Refinement]

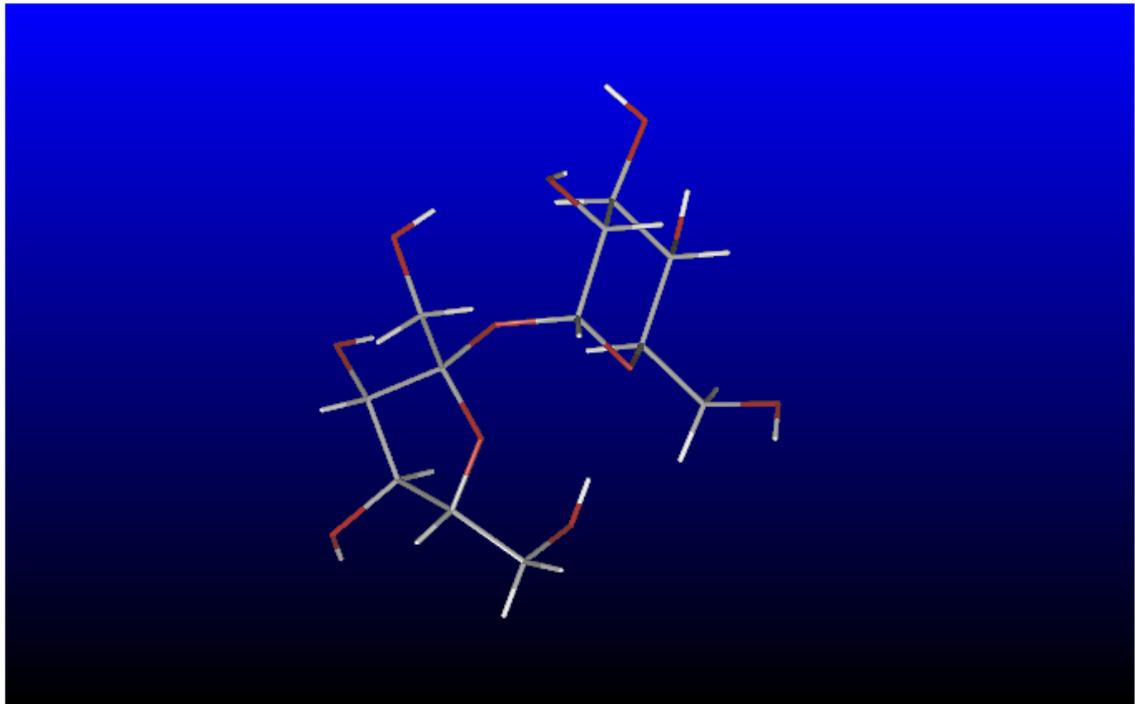
Sample Instrument Windows Help

Setup
Evaluate
Collect
Integrate
Scale
Examine Data
Solve Structure
Refine Structure

Structure Refinement

Instructions Listing Results Structure View

Base: linear_stress_6run



0 No. of QPeaks: 0 5 Zoom

Refresh Files Copy RES to INS Save Files
Refine with XL
Open in XP Open in Olex2 Open in XShell
Open Report

Report
Pilot
Instrument

Hydrogen Bonding

```

Instructions  Listing  Results  Structure View
-----
H12A  2      0.879936   0.724993   0.136234   11.00000   -1.20000
H12B  2      1.073165   0.709329   0.228599   11.00000   -1.20000
AFIX   0
HKLF   4

REM linear_stress_6run in P2(1)
REM R1 = 0.0243 for 3452 Fo > 4sig(Fo) and 0.0248 for all 3505 data
REM 216 parameters refined using 1 restraints

END

WGHT    0.0376    0.1521

REM Instructions for potential hydrogen bonds
EQIV $1 x-1, y, z
HTAB O2 O12_$1
EQIV $2 -x+1, y-1/2, -z+1
HTAB O3 O8_$2
EQIV $3 -x+1, y-1/2, -z
HTAB O4 O6_$3
EQIV $4 x, y-1, z
HTAB O4 O7_$4
EQIV $5 -x+1, y+1/2, -z
HTAB O6 O3_$5
EQIV $6 -x+2, y-1/2, -z+1
HTAB O8 O9_$6
EQIV $7 x+1, y, z
HTAB O9 O11_$7
HTAB O11 O2
HTAB O12 O5
HTAB C1 O6_$5
HTAB C3 O11_$2
HTAB C4 O5_$3
HTAB C6 O12
EQIV $8 -x+1, y+1/2, -z+1
HTAB C8 O2_$8
EQIV $9 -x+2, y+1/2, -z+1
HTAB C10 O8_$9
HTAB C11 O6_$5
EQIV $10 x, y+1, z
HTAB C12 O4_$10
EQIV $11 -x+2, y+1/2, -z
HTAB C12 O6_$11

REM Highest difference peak 0.319, deepest hole -0.179, 1-sigma level 0.042
Q1  1  0.8675  0.5637  0.4638  11.00000  0.05  0.32
Q2  1  0.5377  0.5836  0.3823  11.00000  0.05  0.32
Q3  1  0.5013  0.0911  0.1765  11.00000  0.05  0.29
Q4  1  0.7535  0.1860  0.0860  11.00000  0.05  0.29
Q5  1  0.9127  0.6763  0.2660  11.00000  0.05  0.29

```

- Insert HTAB instruction
- List of possible hydrogen bonds automatically generated
- Review and paste into instructions
- Properly formatted for direct transfer to CIF file

Hydrogen Bonds in .lst File

```

Instructions Listing Results Structure View
n11b      0.9900      109.90      109.90      108.07
          C11 -      O11      C7      H11A

C12 -      Distance      Angles
O12      1.4279 (0.0020)
C10      1.5111 (0.0020) 113.11 (0.12)
H12A     0.9900      108.96      108.96
H12B     0.9900      108.96      108.96      107.76
          C12 -      O12      C10      H12A

Hydrogen bonds with H..A 110 deg.
Appropriate HTAB instructions appended to .res file for future use.
D-H      d(D-H)      d(H..A)      <DHA      d(D..A)      A
O2-H2     0.840      1.989      169.40      2.819      O12 [ x-1, y, z ]
O3-H3     0.840      1.983      176.41      2.822      O8 [ -x+1, y-1/2, -z+1 ]
O4-H4     0.840      2.536      159.80      3.337      O6 [ -x+1, y-1/2, -z ]
O4-H4     0.840      2.355      113.55      2.799      O7 [ x, y-1, z ]
O6-H6     0.840      1.976      164.97      2.796      O3 [ -x+1, y+1/2, -z ]
O8-H8     0.840      1.991      167.77      2.818      O9 [ -x+2, y-1/2, -z+1 ]
O9-H9     0.840      1.864      168.89      2.693      O11 [ x+1, y, z ]
O11-H11   0.840      1.971      155.78      2.759      O2
O12-H12   0.840      1.987      170.60      2.819      O5
C1-H1     1.000      2.319      167.35      3.302      O6 [ -x+1, y+1/2, -z ]
C3-H3A    1.000      2.514      162.54      3.481      O11 [ -x+1, y-1/2, -z+1 ]
C4-H4A    1.000      2.611      130.75      3.351      O5 [ -x+1, y-1/2, -z ]
C6-H6A    0.990      2.538      132.25      3.286      O12
C8-H8A    1.000      2.461      158.86      3.413      O2 [ -x+1, y+1/2, -z+1 ]
C10-H10   1.000      2.640      122.98      3.293      O8 [ -x+2, y+1/2, -z+1 ]
C11-H11B  0.990      2.507      154.62      3.428      O6 [ -x+1, y+1/2, -z ]
C12-H12A  0.990      2.568      125.52      3.245      O4 [ x, y+1, z ]
C12-H12B  0.990      2.512      125.47      3.190      O6 [ -x+2, y+1/2, -z ]

FMAP and GRID set by program

```

Creating Hydrogen Bond Table in CIF file



Report Atoms Bond lengths Bond angles Torsion angles Hydrogen bonds Authors Miscellaneous

```
loop_
  _geom_hbond_atom_site_label_D
  _geom_hbond_atom_site_label_H
  _geom_hbond_atom_site_label_A
  _geom_hbond_site_symmetry_A
  _geom_hbond_distance_DH
  _geom_hbond_distance_HA
  _geom_hbond_distance_DA
  _geom_hbond_angle_DHA
  _geom_hbond_publ_flag
02 H2 012 1_455 0.84(3) 1.99(3) 2.8189(17) 170.(2) ?
03 H3 08 2_646 0.80(3) 2.02(3) 2.8218(16) 175.(2) ?
04 H4 06 2_645 0.75(3) 2.63(3) 3.3387(18) 159.(3) ?
04 H4 07 1_545 0.75(3) 2.38(3) 2.7998(16) 117.(2) ?
06 H6 03 2_655 0.79(3) 2.03(3) 2.7967(18) 166.(2) ?
08 H8 09 2_746 0.79(3) 2.04(3) 2.8195(17) 168.(2) ?
09 H9 011 1_655 0.80(2) 1.89(2) 2.6928(16) 173.(2) ?
011 H11 02 . 0.79(3) 1.99(3) 2.7603(17) 163.(2) ?
012 H12 05 . 0.85(3) 1.98(3) 2.8184(16) 167.(2) ?
C1 H1 06 2_655 0.91(2) 2.40(2) 3.3016(19) 167.8(18) ?
C3 H3A 011 2_646 0.95(2) 2.56(2) 3.4804(19) 162.2(19) ?
C4 H4A 05 2_645 0.97(2) 2.66(2) 3.3511(19) 128.7(18) ?
C6 H6A 012 . 0.95(2) 2.60(2) 3.286(2) 129.9(18) ?
C8 H8A 02 2_656 0.95(2) 2.50(2) 3.4125(18) 161.2(18) ?
C10 H10 08 2_756 0.94(2) 2.69(2) 3.2935(18) 122.5(16) ?
C11 H11B 06 2_655 0.98(2) 2.50(2) 3.4278(19) 156.9(17) ?
C12 H12A 04 1_565 0.99(2) 2.59(2) 3.244(2) 124.2(16) ?
C12 H12B 06 2_755 0.95(2) 2.52(2) 3.1908(19) 128.1(17) ?
```

- CIF H-bonding table
 - "loop_" and headers generated
 - Data fields populated
- All automatic from instructions inserted to .ins file

CIF and SHEXL Files

PLAT005

PLAT005 Type_5 Check for refinement instruction file No 'refinement details' record was found. *Acta Cryst.* requires the inclusion of the last shelxl.res file (in case of a SHELXL or XL refinement) in the CIF embedded between records with semicolons in position 1, preceded by an '[_iucr_refine_instructions_details](#)' record.

Note: SHELXL2013 will automatically include the final .res as an embedded comment with the dataname '[_shelx_res_file](#)'.

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[IUCr Webmaster](#)

- Include .res file and .hkl file to avoid this alert

The 2013 CIF File

- Notable Changes
 - Inclusion of refinement results
 - Inclusion of intensity data
- Benefits
 - Increases experimental transparency
 - SHELX users can easily review instructions, restraints, results
 - Repeat refinement after extracting files from CIF file
- Drawbacks
 - Including files increases complexity of CIF file
 - Including hkl file may increase file size by thousands of lines

```

_shelx_res_file
;
TITL linear_stress_6run in P2(1)
CELL 0.71073 7.69920 8.64700 10.78730 90.0000 102.9858 90.0000
ZERR 2.00 0.00060 0.00060 0.00080 0.0000 0.0019 0.0000
LATT -1
SYMM -X, 0.5+Y, -Z
SFAC C H O
UNIT 24 44 22
REM Solution 1 R1 0.061, Alpha= 0.0028 in P2(1)
REM Flack x= -0.224 ( 0.183 ) from Parsons' quotients
REM C12 O11
REM Instructions for potential hydrogen bonds
EQIV $1 x-1, y, z
HTAB O2 O12_$1
EQIV $2 -x+1, y-1/2, -z+1
HTAB O3 O8_$2
EQIV $3 -x+1, y-1/2, -z
HTAB O4 O6_$3
EQIV $4 x, y-1, z
HTAB O4 O7_$4
EQIV $5 -x+1, y+1/2, -z
HTAB O6 O3_$5
EQIV $6 -x+2, y-1/2, -z+1
HTAB O8 O9_$6
EQIV $7 x+1, y, z
HTAB O9 O11_$7
HTAB O11 O2
HTAB O12 O5
HTAB C1 O6_$5
HTAB C3 O11_$2
HTAB C4 O5_$3
HTAB C6 O12
EQIV $8 -x+1, y+1/2, -z+1
HTAB C8 O2_$8
EQIV $9 -x+2, y+1/2, -z+1
HTAB C10 O8_$9
HTAB C11 O6_$5
EQIV $10 x, y+1, z
HTAB C12 O4_$10
EQIV $11 -x+2, y+1/2, -z
HTAB C12 O6_$11
L.S. 10
acta

```



The 2013 CIF File

- Structure factors incorporated directly to CIF file as data under “_shelx_hkl_file”
- Checksum included at end of list to prevent tampering
- .res and .hkl files can be extracted from the CIF file using “shredCIF”

```
_shelx_hkl_file
;
  2  0  0  94.40  1.94
  2  0  0  99.26  2.11
  2  0  0  95.38  1.91
  2  0  0  96.59  1.89
  2  0  0  96.14  1.91
 -2  0  0  96.95  1.95
 -2  0  0  94.02  1.90
 -2  0  0  98.19  2.11
 -2  0  0  97.78  1.92
 -3  0  0 333.24  6.43
 -3  0  0 335.35  6.56
  3  0  0 331.67  7.17
  3  0  0 331.75  6.60
  3  0  0 339.17  6.42
  3  0  0 327.17  6.55
  3  0  0 317.59  6.60
  3  0  0 337.35  6.42
 -3  0  0 340.45  7.17
 -3  0  0 339.55  6.41
 -4  0  0  46.65  1.06
  4  0  0  46.32  1.08
  4  0  0  45.77  1.00
 -4  0  0  44.78  1.06
 -4  0  0  47.76  1.08
 -4  0  0  45.69  1.05
  4  0  0  46.54  1.08
  4  0  0  47.07  1.00
  0  0  0   0.00  0.00
;
_shelx_hkl_checksum
```

20093

Report Generator and CIF Template



Sample Instrument Report Windows Help

Setup Evaluate Collect Integrate Scale Examine Data Solve Structure Refine Structure Report

Generate Report

Report Atoms Bond lengths Bond angles Torsion angles Hydrogen bonds Authors Miscellaneous

_chemical_formula_moiety	'C12 H22 O11'
_chemical_formula_sum	'C12 H22 O11'
_chemical_formula_iupac	?
_chemical_formula_weight	342.29
_chemical_absolute_configuration	unk
_chemical_melting_point	?
_space_group_crystal_system	monoclinic
_space_group_name_H-M_alt	'P 1 21 1'
_space_group_name_Hall	'P 2yb'
loop_	
_space_group_symop_operation_xyz	
'x, y, z'	
'-x, y+1/2, -z'	
_cell_length_a	7.6992(6)
_cell_length_b	8.6470(6)
_cell_length_c	10.7873(8)
_cell_angle_alpha	90
_cell_angle_beta	102.9858(19)
_cell_angle_gamma	90
_cell_volume	699.80(9)
_cell_formula_units_Z	2
_cell_measurement_reflns_used	9958
_cell_measurement_theta_min	2.9602
_cell_measurement_theta_max	28.3650
_cell_measurement_temperature	100.(2)

Template File:
C:\bn\src\report\acta.cif

Template Sections:

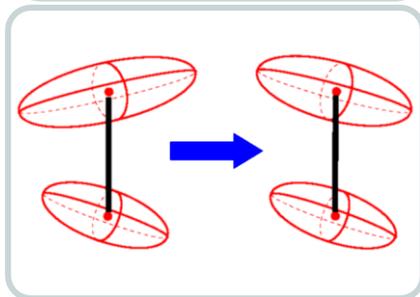
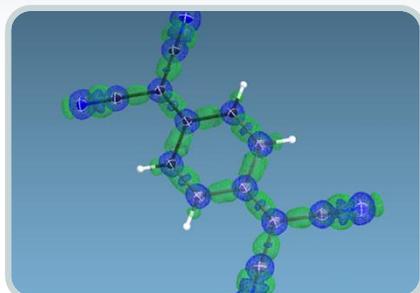
- shex_res_file
- shex_hkl_file
- shex_fab

Save Report...

Open Report in Default Application

12 September 2013 What's New in SHELXL-2013

What's New in SHELXL-2013?



- Increased ability to handle difficult structures
 - RIGU restraint
 - XNPD constraint
 - improved absolute structure determination
- Additional tools for refinement
 - neutron data facilities
 - faster refinement with multi-processor usage
- Improvements in concert with CIF changes
 - SHELX files embedded for re-refinement
 - compliance with core-CIF version 2.4.3.

Conclusion

SHELXL-2013 continues the power and flexibility of SHELXL-97 and its predecessors.

Any questions?

Please type any questions you may have for our speakers in the [Q&A panel](#) and click Send.

How did we do?

When you exit the webinar, please fill out our evaluation [survey](#) to let us know. We appreciate your feedback.

Thank you!





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Webinar	Content
Apr 02, 2013 Fast, intuitive structure Determination II: Crystal Indexing and Data Collection Strategies	Attend this one-hour webinar to learn skills for crystal indexing and data collection. Dr. Bruce Noll will cover the APEX2 tools that can be used to select reflections and sort them into arrays to facilitate indexing routine, twin and modulated structures. He will also examine the diffraction behavior of the sample to determine the best data collection parameters. Register now
Feb 26, 2013 Advanced Crystallography: Data Collection and Data Reduction Techniques for Modulated Structures	Dr. Michael Ruf and Charles Campana give a one-hour presentation demonstrating the steps required to collect and process data for modulated structures. Specific topics include: how to recognize modulated structures; introduction to q-vectors; APEX2 software tools; preparation of files for JANA2006; and examples of the solution of modulated structures from single-crystal diffraction data. View recording Download slides
Dec 05, 2012 PROTEUM2 and Foreign Frame Formats	View this 45-minute webinar to learn about Bruker's PROTEUM software, a suite of crystallographic data integration and reduction tools for macromolecules. Dr. Matt Benning and Dr. Michael Ruf will discuss how to reduce data from various detectors and show results from synchrotron beamlines. View recording Download slides
Nov 13, 2012 Advanced Crystallography: Refinement of Disordered Structures	Dr. Charles Campana gives a one-hour presentation demonstrating the steps required to refine disordered crystal structures. Along with specific examples, topics covered include SHELXL, SHELXTL, free variables, PART n and -n instructions, constraints vs restraints, and advanced refinement instructions. View recording Download slides
Dec 06, 2011 High-Pressure Crystallography	Dr. Francesca Fabbiani of Georg-August-Universität Göttingen takes you through the steps for performing a high-pressure single-crystal X-ray diffraction experiment with a diamond-anvil cell (DAC). You will learn DAC loading techniques and useful strategies for data collection, data reduction, and structure refinement. View recording Download slides
Oct 13, 2011 Advanced Crystallography – Publication of Crystal Structures	Publication of crystal structures is often a researcher's least favorite task. This 90-min webinar will help. It describes the steps required to prepare crystal structures for publication in chemical and crystallographic journals, including collection of high-quality intensity data, common refinement problems, data evaluation, error analysis, preparation of CIF files, and the use of CIF-checking programs. View recording Download slides
Jun 22, 2011 CMOS: The Next Generation in X-ray Detector Technology	CCDs have long been one of the most important detector technologies for X-ray crystallography. Recently, however, new detectors based on CMOS technology have begun to supplant CCDs. View this webinar to learn about Bruker AXS' new X-ray detector based on CMOS technology, the PHOTON 100. View recording Download slides
May 03, 2011 Using APEX2 to Analyze Twinned Crystals	Twinned crystals are common occurrences in many laboratories and pose special challenges in structure solution. View this 76-minute webinar to learn how to visualize, index, integrate, scale, solve and refine structure data from twinned crystals with the APEX2 software suite. (Note: playback begins at 14:00 minutes into the 1:30:20 hour presentation.) View recording Download slides

Accurate 3D structures determined by X-ray diffraction require

A banner image at the top of the slide. On the left, the word "FIRST" is written in large, bold, black letters with white dots between the letters. To its right, a blue bar contains the text "FRONTIERS IN RESEARCH SCIENCE & TECHNOLOGY" in white. Below this bar, the word "newsletter" is written in white on a dark blue background. On the right side of the banner, the Bruker logo is visible, consisting of a blue stylized atom symbol above the word "BRUKER" in bold black letters. The background of the banner shows various scientific images: a circuit board, a test tube with a blue liquid being poured, and a pile of small brown particles.

FiRST

FRONTIERS IN RESEARCH SCIENCE & TECHNOLOGY

newsletter



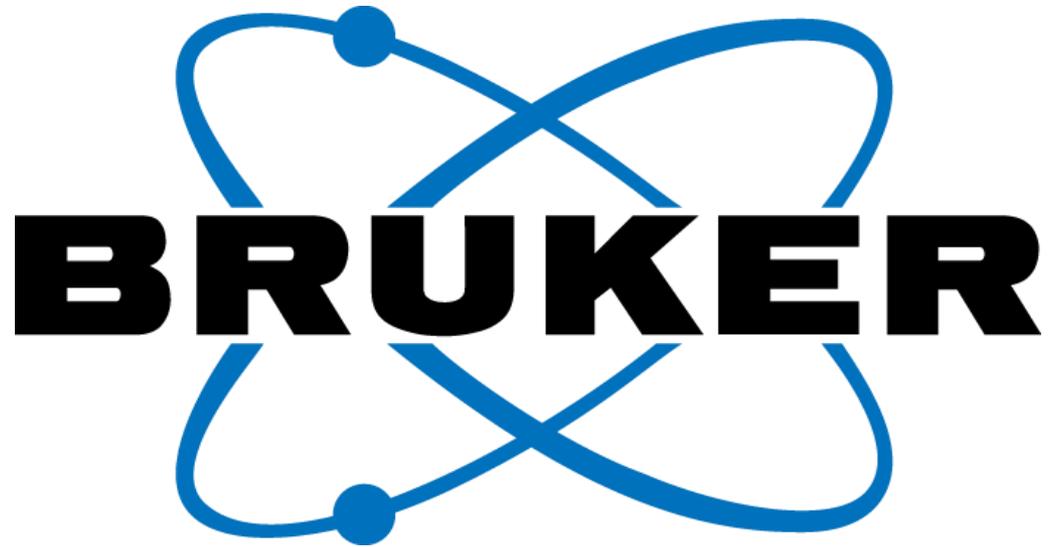
■ ISSUE

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